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NEW DITERPENES FROM SALVIA_TEXANA. CHEMICAL AND BIOGENETIC ASPECTS

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<u>Abstract</u> - Two new diterpenes, the guinone methide 5,6-dehydro- 2α ,7dihydroxytaxodone (1) and 2α -hydroxysalvicanaric acid (5b) were isolated from the roots of <u>Salvia texana</u> Torr Their structure was established by spectroscopic and chemical means. Transformation of 1 into 5b points to 1 being an intermediate in the biogenesis of 5b

A member of the Labiatae family. the genus Salvia consists of some five hundred species found worldwide Since ancient times, many species of this genus have been credited with medicinal properties", and, therefore, reward investigation As part of an intensive study of the chemical composition of the flora used in Latin American popular medicine. following the recent isolation³ of four diterpene methylene guinones, new 6,7-dehydrosalviol and secoditerpenes⁴ from the roots of <u>Salvia_texana</u> Torr. collected two 1 n are now reporting the isolation and structure determination Mexico we of two minor constituents. a guinone methide. 1. and a hemiacetal diterpenic acid, **5b**, as well as touching on some of the chemical and biogenetic aspects involved

The structure of 1 was established as $5,6-dehydro-2\alpha,7-dihydroxytaxodone$ from the following Low resolution m.s showed the molecular ion M^* at m/z346 (molecular formula CzoHzeOs by h r m s) Their spectrum had bands phenols and alcohols (3580, 3520 cm⁻¹) and for a methylene for quinone grouping (1600 and 1570 cm^{-1}), which was confirmed by the n v spectrum (λmax 345, 290 and 255 nm) In the *H n m.r spectrum. signals for an isopropyl group on an aromatic ring and three angular methyls were observed In the low-field region of the spectrum one proton at 7 14. δ interchangeable with deuterium oxide, could be assigned to the phenolic hydroxy group on C-11 while only one proton of the quinone methide system,

Its low chemical shift (6 7 72) indicated the the 14-H. was observed presence of a coplanar hydroxy group on C-7, which was corroborated when the 14-H signal appeared at δ 8.25 when the spectrum was run in py-ds⁵ The CDCl₃ spectrum showed the proton geminal to the alcohol group as a very broad multiplet centred at δ 4.45; in this type of difference, the breadth and multiplicity of this signal is compatible only with a 2-H β and the stereochemistry of the alcohol group was thus determined as 2α . No signals were observed for the $5-H^3$ or for protons allylic to any unsaturated function so the remaining oxygen atom must be part of a Ca-Ca enolic system By means of double resonance and n.O e. difference experiments and a comparison of the ¹H n.m.r. spectra of 1 taken in CDC1₃₅ and $py-d^{35}$, the chemical shifts of all the protons and methyl groups could be ascertained All the above data are in agreement with the structure given for compound 1





AcO

AcO



OAc



(4)

(2)

(3)

ÓН

SCHEME 1

Chemical proof for structure 1 for this new compound was forthcoming as treatment of 1 with methyl iodide and potassium carbonate in acetone gave the dimethylether derivative 2 (Scheme 1) which, in its low resolution m s. showed the molecular ion M^* at m/z 374 (molecular formula $C_{22}H_{30}O_5$ by The 1 r spectrum had bands for enols and alcohols (3590 and hrms)

3370 cm⁻¹) and for a conjugated carbonyl (1620 cm⁻¹) The H nmr spectrum, in addition to the signals for methyl groups and for the 2-HB as a multiplet centred at δ 4 43. showed two methoxy groups as three-proton singlets at δ 3.84 and 3.92. one proton singlet at δ 7.10. interchangeable with deuterium oxide. assigned to 6-OH and the aromatic proton 14-H as a singlet at δ 7 90. a chemical shift characteristic of this type of diterpene when there is a carbonyl group on C-74.7 These data agree with structure 2 which was formed from 1 by tautomerization in basic medium followed by methylation of the phenol groups Treatment of 1 with acetic anhydride 1 n pyridine gave a mixture of two products which were separated by preparative t l c on silica gel The major product was **3** (Scheme 1), low resolution m 5 showed the molecular ion [M⁺] at m/z 472 (molecular formula $C_{2+H_{32}O_{43}}$ by hrms) In its ¹H n m.r spectrum. signals appeared for one aliphatic and two aromatic acetates, the 14-H aromatic proton at δ 8 16, consonant with a ketone group on C-7, and a proton singlet interchangeable with $D_{2}O$ at δ 7 05 assignable to 6-OH The minor product. compound 4. (Scheme 1), had the molecular ion $[M^+]$ at m/z 514 (molecular formula C₂₀H₃₀O₂₀ by h r m s) In the ¹H n m r . signals appeared for one alighting and three aromatic acetates In the low-field region of the spectrum, only the H-14 aromatic proton was observed. as a singlet at δ 8 11 These data are in accordance with structures 3 and 4 for the acetylderivatives of 1.

 2α -Hydroxysalvicanaric acid (**5b**) was isolated as a foam that could not be crystallized Hrms gave the molecular formula. CieHaoOm Bands for acid and aromatic groups and a conjugated carbonyl (1685 cm^{-1}) were seen 1n In the 'H n m r spectrum in py-ds, signals for an isopropyl group on 1 r an aromatic ring and three angular methyls were observed In the low-field region of the spectrum one proton at δ 6 53. interchangeable with deuterium was assigned to a phenol group and a one-proton singlet at 6 7 75 to oxide. the aromatic 14-H The proton geminal to the 2α -hydroxy group appeared as a very broad multiplet centred at δ 4 47

Treatment of **5b** with diazomethane and then with acetic anhydride and ethanol (Scheme 2) gave a mixture of the methyl ester **7** and the acetoxy methyl ester **8**

The main difference between the "H n m r spectra of 7 (CzoHzeOs by and **5b** was the appearance of a carbomethoxy group hrms) ðs a threeproton singlet at δ 3.86 There were signals for only one aromatic acetate group, as a three-proton singlet at δ 2 32, in the 'H nmr spectrum of $8 (C_{22}H_{30}O_7 by h r m s)$ Treatment of **5b** with methyl 10d1de and potassium carbonate in acetone (Scheme 2) gave the dimethoxy derivative 6 (C₂₁H₃₀O₆ by h r m.s.) the ¹H n.m.r of which is very close to that of 7 with one extra signal for a methoxy group as a three-proton singlet at δ 3 98



When **5b** was treated with acetic anhydride in pyridine for five hours (Equation 1), t 1 c. detected a mixture of two products, 9 and 10 When the reaction was prolonged overnight, only 10 was obtained 1 r The spectrum of 9 (CasHsoOm by h.r.m.s.) showed bands for alcohol, carboxylic acid and ester groups In the ¹H n.m r spectrum, signals were observed for one aromatic and one alignatic acetate group as singlets at δ 2 34 and δ 1 96, respectively The 2-HB appeared as a broad multiplet centred at δ



5 13 No geminal proton was observed for the non-acetylated alcohol group, demonstrating its tertiary nature. The hemiacetalic nature of the B ring and consequently of the remaining oxygen atom in the molecule could be deduced

from the ¹³C n m r spectrum of **5b** where a singlet at 114.9 ppm must be attributed to C~5 This spectrum is very similar to that of salvicanaric acid (**5a**), previously isolated from <u>Salvia canariensis</u>, except for the chemical shifts of the C-1, C-2 (in **5b**, a doublet at 63.1 ppm) and C-3 These data all accord with the structure of 2α -hydroxysalvicanaric acid for **5b**.

Chemical proof for structure 5b was provided by 10 being formed when 5b was ¹H n m r. of 10 showed it to be a mixture of 10a and 10b which acetylated behave as one on g c $(C_{22}H_{32}O_{7} by h r m s.)$ The ¹H n.m.r spectra of the two isomers showed signals for an aliphatic acetate and two aromatic acetates in each The 1 r. of the mixture did not have any carboxylic acid absorptions but did have a signal for an alcohol group as well as a lactone 1750 cm⁻¹ The alcohol group had to be tertiary, since it did not at acetylate under the usual conditions and no protons geminal to the hydroxy were observed in ¹H n m.r. The chemical shift of the aromatic 14-H to δ 8 03 (in 10a) and to δ 8 08 (in 10b) places the lactone group carbonyl on C-7 All these data indicate a lactol structure for 10, which must have been formed by the opening of **5b** to its ketophenol tautomer and subsequent acid group attack on the ketone.

Reflux of 10 with acetic anhydride in pyridine (Equation 2) gave the tetracetoxy derivative 11 ($C_{23}H_{34}O_{10}$ by h r.m s) The ¹H n m r spectrum had



signals for two aliphatic and two aromatic acetate groups; the aromatic 14-H appeared as a singlet at δ 8 02. When 10 was treated with diazomethane 1n ether, the ketotriacetoxymethyl ester 12 was obtained as sole product Compound 12 ($C_{2a}H_{3a}O_{9}$ by h.r.m.s) had 1 r signals characteristic of a cyclohexanone and conjugated carboxylate groups In the ¹H n.m r spectrum signals were seen for one aliphatic and two aromatic acetate groups, and also for a carbomethoxy group as a three-proton singlet at δ 3 83 A11 these data accord with the structure 12 which must have been formed via the tautomeric keto-acids of 10

Comparison of the ¹H n.m.r. spectra of **10a** + **10b** run in CDCl₃ and py-d₅ shows that the minor isomer is **10b** with a 58-OH. In the spectrum run in CDCl₃, the 19-Me and 20-Me appear at δ 1 20 and 1.30, respectively, and when taken in py-d₅, at δ 1.44 and 1.59 These shifts were not observed for the major isomer, **10a**

The <u>cis</u> nature of the A/B ring junction in **5b** was established from the following; in the ¹H n.m.r. spectrum of **5b**, the C-20 methyl appears as a three-proton singlet at δ 1.68; when the same spectrum was run in py-d_m it appeared at δ 2 20 in accordance with the <u>cis</u> coplanarity with C_m-OH In a n 0 e difference experiment, irradiation of C-20 methyl produced a strong n 0 e effect on both the H-28 and the C-19 methyl, which agrees with the stereochemistry shown for **5b**

In a previous paper[®], we postulated the biogenetic origin of salvicanaric acid (5a) from the biosynthetic oxidation of demethylcryptojaponol To



confirm this, 1 was auto-oxidated with molecular oxygen in t-butanol and Under potassium t-butoxide, and an intractable mixture was obtained identical conditions, the dimethoxy derivative 2 (Equation 3) gave a product which showed one spot on t l.c but in ¹H n m r proved to consist of а mixture of the lactols 13a and 13b ($C_{21}H_{30}O_4$ by h r.m s) with 13b as the Comparison of the ¹H n m r spectra of **13b** and **13a** taken in major product CDCl₃ and py-d₅ show a great downfield chemical shift of the 18-, 19and 20-Me for the major isomer which must. therefore, have an A/B cis junction An unusual cis ring junction in a similar lactol has already been The aromatic 14-H protons appeared at δ 7 79 in 13b and at δ described? 7 80 in 13a If the phenol groups are free, cyclization after auto-oxidation When 1. dissolved in n-hex-EtOAc, was irradiated should give **5b** (Scheme 3) with u v light (240 nm) for 1/2 hr, two spots were observed on t 1 c the less polar being unreacted starting material When the solution was left to stand, the more polar product slowly evolved to give 5b Heating the



(5b)

solution accelerated the process. Scheme 4 outlines the proposed mechanism via oxygen singlet activation of the substrate as autosensitizer. No intermediate could be isolated.







SCHEME 4

Compound 1 was stable after 24 hr when a solution in n-hex-EtOAc was stirred under oxygen atmosphere, alone or with silica gel added. It was also stable under silica gel chromatography. The above reactions suggest that **5b** may have been formed by the biosynthetic oxidation of 1.

EXPERIMENTAL

Gas chromatography was carried out on a Perkin-Elmer 900 with a flame ionization detector ³H and ³³C n.m r. were collected on Bruker AC80 and WP-200SY (200MHz) spectrometers with $CDCl_{s}$. Py-ds or $CD_{s}OD$ as solvents and TMS as internal standard The 1 r spectra were taken on a Perkin-Elmer 681 spectrophotometer with sodium chloride cells (0.1 mm) and CHCls as solvent spectra were recorded on a Perkin-Elmer 550SE spectrophotometer with UV quartz cells (1 and 5 mm) and EtOH as solvent. Mass spectra and accurate mass measurements were determined on a VG-Micromass ZAB-2F at an lonizing potential of 70 Kev. Dry column chromatography used silica gel (0 05 - 0 2mm) Preparative t.l.c was developed on precoated Schleicher & Schüll foils, F-1500/LS 254 Voucher plant specimens are lodged with the Herbarium of the Dept of Botany, Instituto Tecnológico y de Estudios Superiores de Monterrey, Monterrey, Mexico

The finely-cut roots of <u>Salvia texana</u> Torr (3 kg) were extracted with cold MeOH (5 1) Filtration and evaporation of the solvent with a rotavapor <u>in</u> <u>vacuo</u> gave a reddish-brown extract (48 g) which was chromatographed on Sephadex using a mixture of n-hexane-CHCl₃-MeOH (1 1 2) as solvent 500 ml fractions were collected Only Fractions 40-58 were studied After repeated chromatography on silica gel using mixtures of n-hexane-EtOAc as solvent, the following compounds were isolated -

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Treatment of 1 with Methyl Iodide in Acetone i (10 mg), dissolved in dry acetone (7 ml), was chilled to 5° and potassium carbonate (400 mg) and methyl iodide (1 ml) were then added The reaction mixture was stirred at filtered and taken to dryness and after preparative room temp for 8 hr, chromatography gave 2. (Found: M+, 374 2102 CzzHsoOs requires M 374 2111), Chromatography gave 2. (Found: M^{-} , 3/4 2102 Cz2nsoos requires m 5/4 2111), max (CHCls) cm⁻¹, 3590m, 3370m, 3000w, 2960s, 2920s, 1620s, 1595m, 1470m, 1425w, 1330vs, 1270w and 1040s; max (EtOH) nm, 352, 325, 283 and 255, δ^{44} (200 MHz, CDCls, TMS) 1 24, 1 27 (each 3H. d, J 7Hz, 16-Me and 17-Me), 1 47 (3H, s, 19-Me), 1 54 (3H, s, 20-Me), 1 56 (3H, s, 18-Me), 1.60 (2H, m, 1-Ha and 3-Ha), 2 29 (1H, dd, J 7, 14Hz, 3-Ha), 3 30 (1H, hept, J 7Hz, 15-H), 3 56 (1H, dd, J 7.5, 16Hz, 1-Ha), 3 84, 3.92 (each 3H, s, 2x0Me), 4 43 (1H, br m, W_{1/2} 33Hz, 2-Ha), 7 10 (1H, s, 6-0H), 7 90 (1H, s, 14-H); m/z (e 1) 374 (M* 17%) 355(12) 341(19) 331(8) 325(12) 313(17), 303(12) 374 (M+, 17%), 356(12), 341(19), 331(8), 325(12), 318(32), 313(17), 303(12), 287(100) and 275(99) Treatment of 1 (3 mg) with Ac₂O (0 1 ml) in py (0 2 ml) <u>Acetylation of 1</u> at room temp after 4 hr gave a mixture of **3** and **4**, separated by preparative plate chromatography using benzene-EtOAc (5 5) as eluent **3** was the major product (Found M⁺, 472 2086. C₂₆H₃₂O_B requires M 472 2097), δ^{H} (200 MHz, CDCl₃, TMS) 1 24, 1 28 (each 3H, d, J 7Hz, 16-Me and 17-Me), 1 48 (3H, s, 19-Me), 1 51 (3H, s, 20-Me), 1 54 (3H, s, 18-Me), 2 01 (3H, s, 2-OAc_a), 2 32, 2 39 (each 3H, s, 2xAr-OAc), 2 94 (1H, hept, J 7Hz, 15-H), 5 35 (1H, br m, W_{1/2} 33Hz, 2-H_a), 7 05 (1H, s, 6-O<u>H</u>), 8 16 (1H, s, 14-H), m/z (e 1) 472 (M⁺, 4%), 430(2), 412(16), 402(2), 370(82), 328(100) and 300(8) The minor product A: (found: M⁺ 514 2169, C₂-H₂O₂, C₂-H₂O₂, δ^{H} 4/2 (M^{*}, 4%), 430(2), 412(10), 402(2), 570(62), 528(100) and 500(6) The minor product 4 (found M⁺, 514 2169 $C_{2BH_{34}O_{7}}$ requires M 514 2202), 6⁴⁴ (200 MHz, CDC1₃, TMS) 1 24, 1 25 (each 3H, d, J 7Hz, 16-Me and 17-Me), 1 43 (3H, s, 19-Me), 1 52 (3H, s, 20-Me), 1 57 (3H, s, 18-Me), 2 00 (3H, s, 2-OAc₃), 2 31, 2 36 (each 3H, s, 2xAr-OAc), 2 34 (3H, s, 6-OAc), 3 00 (1H, hept, J 7Hz, 15-H), 5 31 (1H, br m, $W_{1/2}$ 33Hz, 2-H_n), 8 11 (1H, s, 14-H), m/z (e 1) 514 (M⁺, 4%), 472(6), 454(17), 412(68), 388(3), 370(100), 328(85) and 272(20) 2α -Hydroxysalvicanaric Acid (Sb) was isolated as a foam that would not

crystallize (Found M⁺, 350 1732 C₁+H₂₄O₄ requires M 350 1735), max (film) cm⁻¹, 3700-2300vs, 1685vs, 1620m, 1590w, 1470w, 1425vs, 1370m, 1238s, 1090m, 1045m, 1015m, 960w and 910w, max (EtOH) nm, 330, 290 and 250; 6^H (200 MHz, CDCl₃, TMS) 1 23, 1 26 (each 3H, d, J 7Hz, 16-Me and 17-Me), 1 25, 1 27 (each 3H, s, 18-Me, 19-Me), 1 68 (3H, s, 20-Me), 1 80 (1H, dq, 1-H_a), 2 80 (1H, dq, 1-H_a), 3 25 (1H, hept, J 7Hz, 15-H), 4 07 (1H, br m, W_{1/2} 33Hz, 2-H_a), 7 45 (1H, s, 14-H), $\delta^{\rm m}$ (200 MHz, Py-d₅, TMS), 1 24 (3H, s, 18-Me), 1 30, 1 33 (each 3H, d, J 7Hz, 16-Me and 17-Me), 1 49 (3H, s, 19-Me), 2 07 (1H, dq, 1-H_a), 2 24 (3H, s, 20-Me), 3 68 (1H, hept, J 7Hz, 15-H), 3 74 (1H, dq, 1-H_a), 4 52 (1H, br m, W_{1/2} 33Hz, 2-H_a), 7 84 (1H, s, 14-H), $\delta^{\rm m}$ (200 MHz, Py-d₅, TMS), 1 24, (3H, s, 19-Me), 2 07 (1H, dq, 1-H_a), 4 52 (1H, br m, W_{1/2} 33Hz, 2-H_a), 7 84 (1H, s, 14-H), $\delta^{\rm m}$ (200 MHz, Py-d₅+D₂O, TMS), 1 25, 1 28 (each 3H, d, J 7Hz, 16-Me and 17-Me), 1 32 (3H, s, 18-Me), 1 43 (3H, s, 19-Me), 2 10 (1H, dq, 1-H_a), 2 23 (3H, s, 20-Me), 3 62 (1H, hept, J 7Hz, 15-H), 3 78 (1H, dq, 1-H_a), 4 50 (1H, br m, W_{1/2} 33Hz, 2-H_a), 7 78 (1H, s, 14-H), $\delta^{\rm m}$ (200 MHz, CD₃OD, TMS), 1 19 (6H, d, J 7Hz, 16-Me, 17-Me) 1 28 (3H, s, 18-Me) 1 59 (3H, s, 19-Me), 1 90 (3H, s, 20-Me), 2 68 (1H, dq, 1-H_a), 3 85 (1H, br m, W_{1/2} 33Hz, 2-H_a), 6 90 (1H, s, 14-H), $\delta_{\rm m}$ (50MHz, Py-d₅), 18 9 (q, C-20), 22 97 (q, C-16), 23 03 (q, C-17), 25 8 (q, C-18), 27 3 (q, C-19), 27 4 (d, C-15) 39 6 (s, C-4), 48 1 (t, C-3), 49 2 (t, C-1), 52 4 (s, C-10), 63 1 (d, C-2), 114 9 (s, C-5), 120 7 (s C-9), 122 0 (d C-14), 135 2 (s, C-8), 139 3 (s, C-11), 144 5 (s, C-13), 304(2), 288(5), 273(3), 250(38), 237(28), 219(15), 189(11), 161(8), 119(16) and 55(100)

5b (8 7 mg), dissolved in ether (5 ml), Treatment of **5b** with Diazomethane treated with an ether soln of CH_2N_2 (2ml. 0 4M) After 20 min the wag reaction was halted by the addition of Ac20 and the excess eliminated by Evaporation of the soln in rotavapor in vacuo gave a addition of EtOH mixture of 7 and 8. separated by preparative t 1 c 7: (found M+ . 364 1903 CzoHzeOz requires M 364 1920), max (CHClz) cm⁻¹, 3670m, 3590m, 3550m, 3000m, 2950m, 2920m, 1710vs, 1610m, 1600m, 1440m, 1420s, 1370m, 1210vs, 1100s and 1040m, δ^H (200 MHz, CDC1₃, TMS) 1 23, 1 27 (each 1335m. 3H, d, J 7Hz, 16-Me and 17-Me), 1 24, 1 25 (each 3H, s, 18-Me, 19-Me), 1 60 (3H, s, 20-Me), 1 76 (1H, dq, 1-H_a), 2 72 (1H, dq, 1-H_a), 3 25 (1H, hept, J 7Hz, 15-H), 3 86 (3H, s, -COOMe), 4 03 (1H, br m, $W_{1/2}$ 33Hz, 2-H_a), 5 30 (1H, br s, ArOH), 7 29 (1H, s, 14-H), m/z (e 1) 364 (M⁺, 18%), 346(16), 333(3), 315(12), 305(1), 302(16), 287(10), 264(100), 249(37) and 221(38) B (found M⁺, 406 1998 C₂₂H₃₀O₇ requires M 406 2005), S^H (200 MHz, CDCl₃, TMS) 1 19, 1 21 (each 3H, s, 18-Me, 19-Me), 1.21 (6H, d, J 7Hz, 16-Me, 17-Me), 1 60 (3H, s, 20-Me), 1 72 (1H, dq, 1-H_a), 2 32 (3H, s, Ar-OAc), 2 72 dq, 1-H_a), 3 10 (1H, hept, J 7Hz, 15-H), 3 59 (3H, s, -COO<u>Me</u>), 4 02 (1H. br m, $W_{1/2}$ 33Hz, 2-HB), 7 26 (1H, s, 14-H), m/z (e 1) 406 (M^+ , 11%), (1H. 364(12), 346(1), 333(1), 315(2), 306(53), 302(1), 287(5), 278(10) and 264(100)

<u>Acetylation of 5b</u> Treatment of 5b (10 mg) with $Ac_{2}O$ (0 3 ml) in py (0 6 ml) at room temp for 5 hr gave 7 + 10 If the reaction was left to stand overnight only 10 was obtained

9 (found M⁺, 434 1943 C_{2xH}_{xo}O₂ requires M 434 1946), max (CHCl₃) cm⁻¹, 3670w, 3020w, 2860m, 2820w, 1750m, 1735vs, 1420s, 1370s, 1250vs, 1190s, 1095m and 1020m, 6⁴⁴ (200 MHz, CDCl₃, TMS) 1 20 (6H, s, 18-Me, 19-Me), 1 22, 1 26 (each 3H, d, J 7Hz, 16-Me, 17-Me), 1 68 (3H, br s, 20-Me), 1 96 (3H, s, 2-OAc_x), 2 34 (3H, s, Ar-OAc), 2 70 (1H, dq, 1-H_A), 3 06 (1H, hept, J 7Hz, 15-H), 5 13 (1H, br m, W_x_x_33Hz, 2-H_a), 7 42 (1H, s, 14-H), 6⁴⁴ (200 MHz, Py-d₅, TMS) 1 15, 1 17 (each 3H, d, J 7Hz, 16-Me, 17-Me), 1 46, 1 59 (each 3H, s, 18-Me, 19-Me), 1 97 (3H, s, 2-OAc_x), 2 14 (3H, s, 20-Me), 2 37 (3H, s, Ar-OAc), 3 12 (1H, hept, J 7Hz, 15-H), 3 31 (1H, dq, 1-H_b), 5 30 (1H, br m, W_x_x_33Hz, 2-H_a), 7 67 (1H, s, 14-H), m/z (e 1) 434 (M⁺, 1%), 392(3), 322(3), 314(2), 292(17), 279(10), 265(4), 250(25), 167(20) and 149(100) **10** (found M⁺, 476 2059 C_{2xH}_{xa}O₂ requires M 476 2072), max (CHCl₃) cm⁻¹, 3660m, 3580m, 3000s, 2960s, 2920s, 1780vs, 1750vs, 1620w, 1380m, 1325m, 1250s, 1195s, 1180m, 1100m and 1040s, δ^{H} (200 MHz, CDL₃, TMS) **10a** 1 20 (3H, s, 20-Me), 2 05 (3H, s, 2-OAc_a), 2 27, 2 31 (each 3H, s, 2vAr-OAc), 2 67 (1H, dd, 1-H_a), 2 95 (1H, hept, J 7Hz, 15-H), 3 26 (1H, br s, 5-OH), 5 24 (1H, br m, W_x_x_33Hz, 2-H_a), 8 03 (1H, s, 14-H), **10b** 1 20 (3H, s, 18-Me), 1 20 (3H, s, 19-Me), 1 22 (6H, d, J 7Hz, 16-Me, 17-Me), 1 30 (3H, s, 20-Me), 1 97 (3H, s, 2-OAc_a), 2 31, 2 32 (each 3H, s, 2vAr-OAc), 2 67 (1H, dd, 1-H_a), 2 95 (1H, hept, J 7Hz, 15-H), 3 13 (1H, dq, 1-H_b), 5 42 (1H, br m, W_x_x_33Hz, 2-H_a), 8 08 (1H, s, 14-H), 6⁴⁴ (200 MHz, S-DH), 5 42 (1H, br m, W_x_x_33Hz, 2-H_a), 8 08 (1H, s, 14-H), 10b 1 00 (3H, s, 20-Me), 1 97 (3H, s, 2-OAc_a), 2 31, 2 32 (each 3H, s, 2vAr-OAc), 2 67 (1H, dd, 1-H_a), 2 97 (1H, hept, J 7Hz, 15-H), 3 13 (1H, dq, 1-H_b), 5 42 (1H, br m, W_x_x_33Hz, 2-H_a), 8 08 (1H, s, 14-H), 6⁴⁴ (200 MHz, S-DH), 5 42 (1H, br m, W_x_x_33Hz, 2-H_a), 8 28 (1H, s, 14-H), 10b 1 06, 1 10 (each 3H, d, J 7Hz, 16-Me, 17-Me), 1 36 (

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